IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Atty. Docket: TOVEY5A

In re Application of:) Conf. No.: 1014
)
Michael G. TOVEY et al.) Art Unit: 1636
)
Appln. No.: 10/677,777) Examiner: L. Mitchell
)
Filed: October 3, 2003) Washington, D.C.
)
For: GENE REPORTER ASSAY, AND)
CELLS FOR DETERMINING THE)
PRESENCE AND/OR THE …)

DECLARATION UNDER 37 CFR §1.132

Honorable Commissioner for Patents
U.S. Patent and Trademark Office
Customer Service Window
Randolph Building, Mail Stop **Amendment**
401 Dulany Street
Alexandria, VA 22314

Sir:

I, Michael G. TOVEY, hereby declare and state as follows:

I am a coinventor of the above-identified application and my educational and professional experience is presented in the curriculum vitae attached hereto.

I understand that the cell line and kit claims in the above-identified application are being rejected because the specification is considered by the examiner to lack enablement for any cell line other than a promonocytic cell line. The examiner has stated the specification discloses that a treated human promonocytic cell line will have a shelf life of about 24 hours and that the time of exposure to vinblastin is variable, but also that the formulation of vinblastin is important for the desired outcome. It is the examiner's position that applicants do not provide any further specific guidance regarding how much vinblastin or cisplatin or 5-fluorouracil should be provided to any other cell type at any other concentration.

The experiments presented below demonstrate that a variety of human cell types (promonocytic, myeloid, T-cell lymphoma, breast adenocarcina) and non-human cells (mouse lymphoma), all treated with the same concentration of vinblastin and conditions before being cryopreserved in ampoules/vials as taught in the above-identified application on pages 47 and 48 of the specification, are all capable of maintaining signal transduction activity for at least 24 hours after being thawed (and before undergoing cellular death in no more than 30 days at a temperature above freezing). These experiments were conducted under my direct supervision and I can attest of my own personal knowledge that all the results reported herein are true and accurate.

The following experimental conditions were optimized for PIL5 cells, so as to obtain induction of apoptosis in 100% of the cell population while maintaining fully functional signal transduction for at least 24 hours after thawing vinblastin-treated cells. These same optimal conditions were then applied to the other cell lines/types without further experimentation and were found to induce apoptosis in 100% of each of the cell populations while maintaining fully functional signal transduction for at least 24 hours after thawing vinblastin-treated cells.

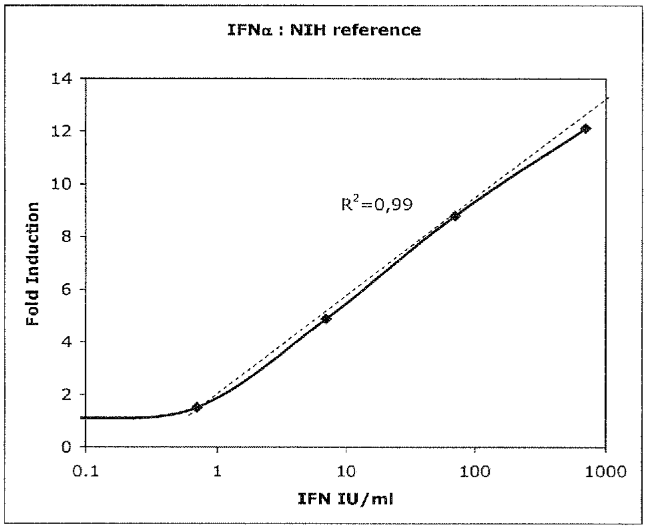
Human type I Interferons (Interferon alpha & Interferon Beta)

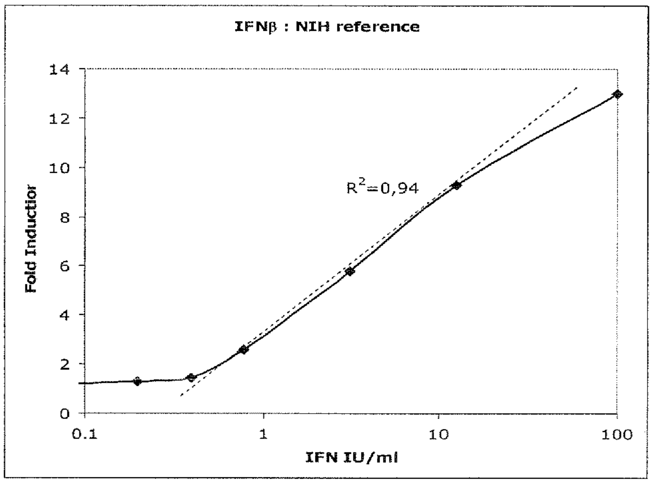
PIL5 cells, a clone of the human pro-monocytic cell line U937 transfected with the ISRE from the ISG15 gene controlling a SV40 minimal promoter regulating the transcription of the fire-fly luciferase reporter gene, were seeded at a concentration of 0.5 to 1 x 10^5 cells/ml in RPMI 1640 medium supplemented with 10% fetal bovine serum (FBS) and harvested when the cells had reached a concentration of 2 to 7 x 10^5 cells/ml. The cells were then treated for 10 minutes at 37°C with a freshly prepared solution of vinblastin at a final concentration of 1.0 microgram/ml RPMI 1640 in medium supplemented with 10% FBS. Vinblastin (commercially available from Eli Lilly under the pre-buffered formulation VELBE) was diluted from a 1 mg/ml stock solution in H_2O.

The PIL5 cells were then centrifuged at 800 x g for 10 minutes at 4°C and washed once with the same volume of RPMI 1640 medium with 10% FBS to remove the vinblastin. The PIL5 cells were then re-suspended at a concentration of 2 x 10^7 cells/ml RPMI 1640 medium with 40% FBS and 2.5% dimethylsulfoxide + 10% glycerol. The cell suspension (1.0 ml) was then dispensed into cryopreservation vials and frozen at -80° C.

Serial dilutions of the human IFN alpha or IFN beta NIH/WHO reference preparation and the samples to be assayed were prepared in a 96-well micro-titer plate (CulturPlate96, Perkin Elmer, Waltham MA) in 50 micro-liters of RPMI 1640 + 1% FBS. A vial of frozen vinblastin-treated cells was thawed in a 37°C water-bath immediately prior to performing the assay, diluted in 9 ml of RPMI 1640 + 1% FBS, and 50 micro-liters of the cell suspension was added to each well of the micro-titer plate and incubated overnight (>16 hours) at 37°C in 5% CO_2 in air. One hundred micro-liters of luciferin containing lysis buffer (SteadyLite Plus HTS, Perkin Elmer, Waltham MA) was added to each well of the micro-titer plate. The luminescence was then determined in a micro-plate reading luminometer (Lumi Count, Packard-Perkin Elmer, Waltham MA) after 5 minutes incubation at room temperature. The standard curves for IFNα and IFNβ

presented below show that the vinblastin-treated cells maintain the desired fully functional signal transduction activity at least 24 hours after thawing.





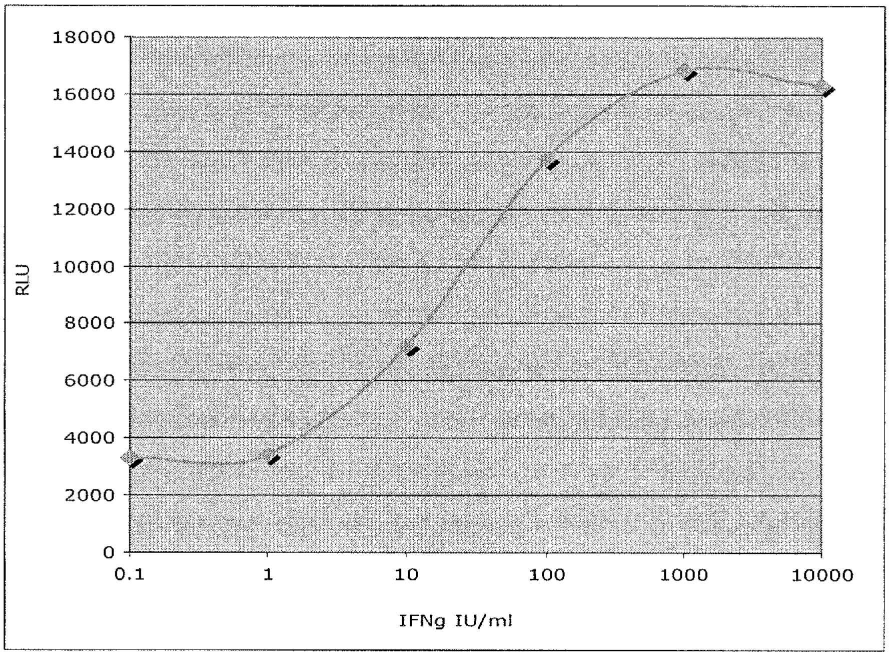
Human Type II Interferon (Interferon Gamma)

UIG2 cells, a clone of the human myeloid cell line U266R transfected with the GAS from the IFR1 gene controlling a SV40 minimal promoter regulating the transcription of the fire-fly luciferase reporter gene, were seeded at a concentration of 0.5 to 1 x 10^5 cells/ml in RPMI 1640 medium supplemented with 10%

FBS and harvested when the cells had reached a concentration of 2 to 7 x 10^5 cells/ml. The cells were then treated for 10 minutes at 37°C with a freshly prepared solution of vinblastin at a final concentration of 1.0 microgram/ml RPMI 1640 in medium supplemented with 10% FBS. Vinblastin (commercially available from Eli Lilly under the pre-buffered formulation VELBE) was diluted from a 1 mg/ml stock solution in H_2O.

The UIG2 cells were then centrifuged at 800 x g for 10 minutes at 4°C and washed once with the same volume of RPMI 1640 medium with 10% FBS to remove the vinblastin. The UIG2 cells were then re-suspended at a concentration of 2 x 10^7 cells/ml RPMI 1640 medium with 40% FBS and 2.5% dimethylsulfoxide + 10% glycerol. The cell suspension (1.0 ml) was then dispensed into cryopreservation vials and frozen at -80° C.

Serial dilutions of the human interferon gamma reference preparation and the samples to be assayed were prepared in a 96-well micro-titer plate (CulturPlate96, Perkin Elmer, Waltham MA) in 50 micro-liters of RPMI 1640 + 1% FBS. A vial of frozen vinblastin-treated cells was thawed in a 37°C water-bath immediately prior to performing the assay diluted in 9 ml of RPMI 1640 + 1% FBS and 50 micro-liters of the cell suspension was added to each well of the micro-titer plate and incubated for 4 to 6 hours at 37°C in 5% CO_2 in air. One hundred micro-liters of luciferin containing lysis buffer (SteadyLite Plus HTS, Perkin Elmer, Waltham MA) was added to each well of the micro-titer plate. The luminescence was then determined in a micro-plate reading luminometer (Lumi Count, Packard-Perkin Elmer, Waltham MA) after 5 minutes incubation at room temperature. The standard curve for IFNγ presented below shows that the vinblastin-treated cells maintain the desired fully functional signal transduction activity at least 24 hours after thawing.

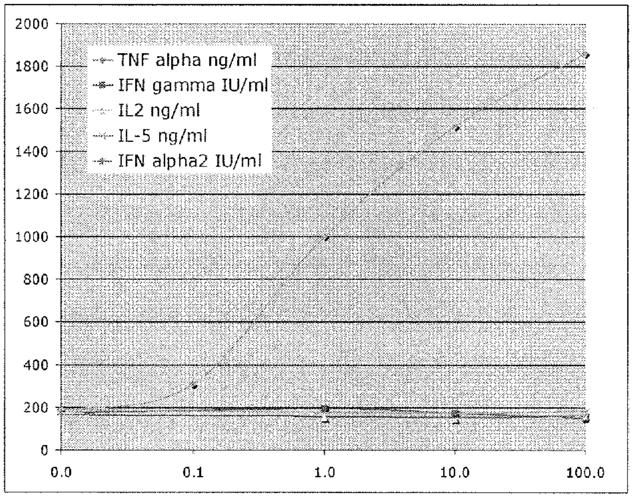


Human TNF-alpha

JUT4 cells, a clone of the human T-cell lymphoma cell line Jurkatt transfected with the 5 x tandem repeat of the canonical NFκB binding site controlling a SV40 minimal promoter regulating the transcription of the fire-fly luciferase reporter gene, were seeded at a concentration of 0.5 to 1 x 10^5 cells/ml in RPMI 1640 medium supplemented with 10% FBS and harvested when the cells had reached a concentration of 2 to 7 x 10^5 cells/ml. The cells were then treated for 10 minutes at 37°C with a freshly prepared solution of vinblastin at a final concentration of 1.0 microgram/ml RPMI 1640 in medium supplemented with 10% FBS. Vinblastin (commercially available from Eli Lilly under the pre-buffered formulation VELBE) was diluted from a 1 mg/ml stock solution in H_2O.

The JUT4 cells were then centrifuged at 800 x g for 10 minutes at 4°C and washed once with the same volume of RPMI 1640 medium with 10% FBS to remove the vinblastin. The JUT4 cells were then re-suspended at a concentration of 2 x 10^7 cells/ml RPMI 1640 medium with 40% FBS and 2.5% dimethylsulfoxide + 10% glycerol. The cell suspension (1.0 ml) was then dispensed into cryopreservation vials and frozen at -80°C.

Serial dilutions of the TNF standard and the samples to be assayed were prepared in a 96-well micro-titer plate (CulturPlate96, Perkin Elmer, Waltham MA) in 50 micro-liters of RPMI 1640 + 1% FBS. A vial of frozen vinblastin-treated cells was thawed in a 37°C water-bath immediately prior to performing the assay diluted in 9 ml of RPMI 1640 + 1% FBS and 50 micro-liters of the cell suspension was added to each well of the micro-titer plate and incubated for 4 to 6 hours at 37°C in 5% CO_2 in air. One hundred micro-liters of luciferin containing lysis buffer (SteadyLite Plus HTS, Perkin Elmer, Waltham MA) was added to each well of the micro-titer plate. The luminescence was then determined in a micro-plate reading luminometer (Lumi Count, Packard-Perkin Elmer, Waltham MA) after 5 minutes incubation at room temperature. The standard curve for TNFα presented below shows that the vinblastin-treated JUT4 cells maintain the desired fully functional signal transduction activity (in response to TNF-α but not to controls IFNγ, IL2, IL5 and IFNα2) at least 24 hours after thawing.

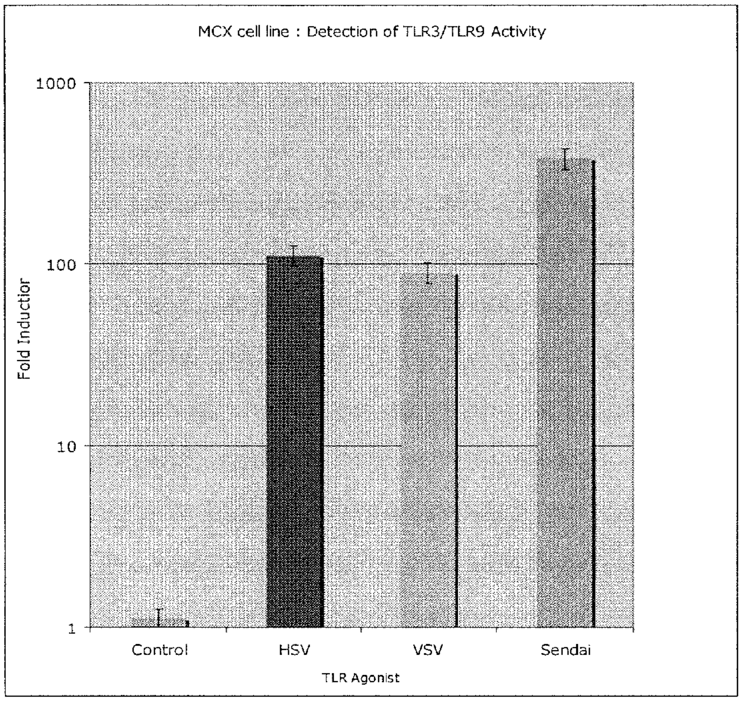


Human Toll Like Receptor Agonists

MCX cells, a clone of the human breast adenocarcinoma cell line MCF7 transfected with 1.2kb of the promoter region of the NOXA gene controlling a SV40 minimal promoter regulating the transcription of the fire-fly luciferase reporter gene were seeded at a concentration of 0.5 to 1 x 10^5 cells/ml in DMEM medium supplemented with 10% FBS and harvested when the cells had reached a concentration of 2 to 7 x 10^5 cells/ml. The cells were then treated for 10 minutes at 37^{o}C with a freshly prepared solution of vinblastin at a final concentration of 1.0 microgram/ml DMEM in medium supplemented with 10% FBS. Vinblastin (commercially available from Eli Lilly under the pre-buffered formulation VELBE) was diluted from a 1 mg/ml stock solution in H_2O.

The MCX cells were then centrifuged at 800 x g for 10 minutes at 4^{o}C and washed once with the same volume of DMEM medium with 10% FBS to remove the vinblastin. The MCX cells were then re-suspended at a concentration of 2 x 10^7 cells/ml DMEM medium with 40% FBS and 2.5% dimethylsulfoxide + 10% glycerol. The cell suspension (1.0 ml) was then dispensed into cryopreservation vials and frozen at -80° C.

Serial dilutions of the TLR agonist or antagonist standard and the samples to be assayed were prepared in a 96-well micro-titer plate (CulturPlate96, Perkin Elmer, Waltham MA) in 50 micro-liters of RPMI 1640 + 1% FBS. A vial of frozen vinblastin-treated cells was thawed in a 37°C water-bath immediately prior to performing the assay diluted in 9 ml of RPMI 1640 + 1% FBS and 50 micro-liters of the cell suspension was added to each well of the micro-titer plate and incubated for 12 to 18 hours at 37°C in 5% CO_2 in air. One hundred micro-liters of luciferin containing lysis buffer (SteadyLite Plus HTS, Perkin Elmer, Waltham MA) was added to each well of the micro-titer plate. The luminescence was then determined in a micro-plate reading luminometer (Lumi Count, Packard-Perkin Elmer, Waltham MA) after 5 minutes incubation at room temperature. Vinblastin-treated MCX cells maintained the desired signal transduction activity from toll-like receptors in response to toll-like receptor agonists HSV, VSV and Sendai viruses at least 24 hours after thawing.

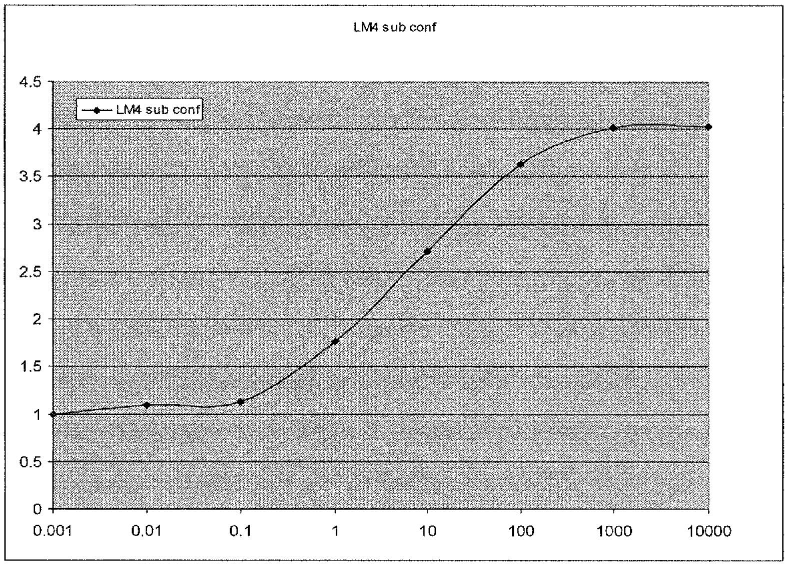


Mouse Type I Interferons

LM4 cells, a clone of the mouse lymphoma L1210 cell line transfected with the ISRE from the ISG15 gene controlling a SV40 minimal promoter regulating the transcription of the firefly luciferase reporter gene were seeded at a concentration of 0.5 to 1 x 10^5 cells/ml in RPMI 1640 medium supplemented with 10% FBS and harvested when the cells had reached a concentration of 2 to 7 x 10^5 cells/ml. The cells were then treated for 10 minutes at 37^{o}C with a freshly prepared solution of vinblastin at a final concentration of 1.0 microgram/ml RPMI 1640 in medium supplemented with 10% FBS. Vinblastin (commercially available from Eli Lilly under the pre-buffered formulation VELBE) was diluted from a 1 mg/ml stock solution in H_2O.

The LM4 cells were then centrifuged at 800 x g for 10 minutes at 4°C and washed once with the same volume of RPMI 1640 medium with 10% FBS to remove the vinblastin. The LM4 cells were then re-suspended at a concentration of 2×10^7 cells/ml RPMI 1640 medium with 40% FBS and 2.5% dimethylsulfoxide + 10% glycerol. The cell suspension (1.0 ml) was then dispensed into cryopreservation vials and frozen at -80° C.

Serial dilutions of the murine IFN alpha or IFN beta reference preparation and the samples to be assayed were prepared in a 96-well micro-titer plate (CulturPlate96, Perkin Elmer, Waltham MA) in 50 micro-liters of RPMI 1640 + 1% FBS. A vial of frozen vinblastin-treated cells was thawed in a 37°C water-bath immediately prior to performing the assay diluted in 9 ml of RPMI 1640 + 1% FBS and 50 micro-liters of the cell suspension was added to each well of the micro-titer plate and incubated overnight (>16 hours) at 37°C in 5% CO_2 in air. One hundred micro-liters of luciferin containing lysis buffer (SteadyLite Plus HTS, Perkin Elmer, Waltham MA) was added to each well of the micro-titer plate. The luminescence was then determined in a micro-plate reading luminometer (Lumi Count, Packard-Perkin Elmer, Waltham MA) after 5 minutes incubation at room temperature. The standard curve for murine IFNα or IFNβ presented below shows that the vinblastin-treated cells maintain the desired fully functional signal transduction activity at least 24 hours after thawing.



In conclusion, irrespective of the cell type or cell line used for treatment with vinblastin, or whether the cells are human or non-human, all cell types/lines tested (which includes a varied sampling of different cell types) using the same vinblastin treatment and cryopreservation conditions taught in the present specification were found to maintain the desired fully functional signal transduction activity in gene reporter assays for at least 24 hours after thawing. There is no reason to doubt that additional further cell types would also work under the same specific vinblastin treatment conditions, which can also certainly be easily modified if needed with only routine experimentation. Indeed, I fully expect that most cell types/lines would maintain the desired signal activity under the same specific optimized vinblastin treatment conditions taught in the present specification, or would require at most a minimal amount of routine experimentation to adjust and optimize.

The undersigned declares further that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

10-16-2007.
Date

Michael Tovey.
Michael G. TOVEY

G:\BN\N\Neux\Tovey5A\pto\2007-10-15Declaration1.132.doc

CURRICULUM VITAE

Michael G. TOVEY

PRESENT AFFILIATION

Director of the Laboratory of Viral Oncology
Chief, CNRS FRE 2942
Institut André Lwoff
7 rue Guy Môquet - BP 8
94801 Villejuif Cedex, France
Tel. (33-1) 49 58 34 23/49 58 34 34.
Fax (33-1) 49 58 34 44
e-mail : tovey@vjf.cnrs.fr

PERSONAL

Birth :	March 8, 1945
Citizenship :	British
Marital Status :	Married, two children

EDUCATION

B.s.C. 1969	Special Microbiology, London University
Ph.D. 1972	Microbiology, London University

RESEARCH AND ACADEMIC APPOINTMENTS

Sept. 1968 - Oct. 1971	Assistant (Department of Microbiology - Prof. S. Pirt) Queen's Elizabeth College, London.
1969 - 1971	Assistant in Biochemistry, Queen's Elizabeth College, London.
	Assistant in Bacteriophage genetics and animal virology, Queen's Elizabeth College, London.
1971 - April 1973	Post-doctoral research fellow of European Molecular Biology Organization at the Institut de Recherches Scientifiques sur le Cancer, Laboratory of Viral Oncology (Dr Ion Gresser), Villejuif, France.
April 1973 - Dec. 1976	Attaché de recherche I.N.S.E.R.M. at the Institut de Recherches Scientifiques sur le Cancer, Villejuif, France.

Dec.1976 - Dec. 1979	Chargé de Recherche I.N.S.E.R.M. at the Institut de Recherches Scientifiques sur le Cancer, Villejuif, France.
Jan.1980 - 1983	Maître de Recherche I.N.S.E.R.M. at the Institut de Recherches Scientifiques sur le Cancer, Villejuif, France.
Jan. 1984 - 1997	Directeur de Recherche 2 I.N.S.E.R.M. at the Institut de Recherches Scientifiques sur le Cancer, Villejuif, France.
Jan. 1998	Directeur de Recherche First class I.N.S.E.R.M. at the Institut de Recherches Scientifiques sur le Cancer, Villejuif, France.
Jan. 1984	Fogarty visiting Professor, Rockefeller University, New York, U.S.A.
1987-1991	Member of the Molecular Biology Commission of INSERM Member of the Stearing Committee of INSERM Molecular Biology Commission
1988-1992	Member of the Scientific Board of Faculty of Medicine, University Paris XI
1989	Founded the European Biotechnology Corporation
Since 1990	Member of the Scientific Board of INSERM Unit 268
January 1995 - present	Director of the Laboratory of Viral Oncology,
1995-2005	Chief, CNRS UPR 9045 at the Institut André Lwoff, Villejuif, France
January 1995 - present	Member of the Board of the André Lwoff Institute,
January 2006 - present	Chief, CNRS FRE 2942 at the Institut André Lwoff, Villejuif, France

MEMBERSHIP OF EDITORIAL BOARDS AND SCIENTIFIC COMMITTEES

- Section Editor Journal of Interferon & Cytokine Research
- Member of the Editorial Board of Oncology Reports
- Member of the Editorial Board of Cellular Pharmacology
- French representative for the ISICR International Council
- Vice-President of the Scientific Board of the Prix Européen de l'Avenir
- Vice-President of the Association of the Institut of Cancer and Immunogenetics, Villejuif, France
- President of the Scientific Board of the Institut of Cancer and Immunogenetics, Villejuif, France
- General Secretary of the Association for Biomedical Research (ANRB)
- Member of the Scientific Board of the International Center for AIDS Research in Africa (CILSA)
- Member of the Scientific Board of Medinvest, Compagnie Financière Edmond de Rothschild

HONORS

1976	Prix ESSEC
1988	Associé au Dr. Gresser, Prix Antoine Lacassagne

PUBLICATIONS

1. Mogensen K.E., Tovey M.G., Mathison G.E. and Pirt S.J.
Induction of mouse interferon in a chemically defined system.
J. gen. Virol., **16** : 111-114, 1972.

2. Tovey M.G., Mathison G.E. and Pirt S.J.
The production of interferon by chemostat cultures of mouse LS-cells grown in chemically-defined, protein-free medium.
J. gen. Virol., **20** : 29-35, 1973.

3. Frayssinet C., Tovey M.G., Gresser I. and Lindahl P.
Inhibitory effect of potent interferon preparations of the regeneration of mouse liver after partial hepatectomy.
Nature, **245** : 146-147, 1973.

4. Gresser I., Bandu M-T., Tovey M.G., Bodo G., Paucker K. and Stewart II W.E.
Inhibitory effect of highly purified interferon preparations on the multiplication of leukemia L1210 cells.
Proc. Soc. Exp. Biol. Med., **142** : 7-10, 1973.

5. Tovey M.G., Begon-Lours J. and Gresser I.
A method for the large scale production of potent interferon preparations.
Proc. Soc. Exp. Biol. Med., **146** : 809-815, 1974.

6. Gresser I., Bandu M-T., Brouty-Boyé D. and Tovey M.G.
Pronounced antiviral activity of human interferon on bovine and porcine cells.
Nature, **251** : 543-545, 1974.

7. Tovey M.G., Brouty-Boyé D. and Gresser I.
Early effect of interferon on mouse leukemia cells cultivated in a chemostat.
Proc. Natl. Acad. Sci. USA, **72** : 2265-2269, 1975.

8. Gresser I., Tovey M.G. and Bourali-Maury C.
Efficacy of exogenous interferon treatment initiated after onset of multiplication of vesicular stomatitis virus in the brains of mice.
J. gen. Virol., **27** : 222-233, 1975.

9. Gresser I., Tovey M.G., Maury C. and Chouroulinkov I.
Lethality of interferon preparations for new-born mice.
Nature, **258** : 76-78, 1975.

10. Tovey M.G. and Brouty-Boyé D.

Characteristics of the chemostat culture of murine leukemia L1210 cells.
Exp. Cell Res., **101** : 346-354, 1976.

11. Tovey M.G. and Brouty-Boyé D.
The cultivation of animal cells in a chemostat.
J. App. Chem. & Biotech., **26** : 345, 1976.

12. Gresser I., Tovey M.G., Bandu M-T., Maury C. and Brouty-Boyé D.
Role of interferon in the pathogenesis of virus diseases in mice as demonstrated by the use of anti-interferon serum. I. Rapid evolution of encephalomyocarditis virus infection.
J. Exp. Med., **144** : 1305-1315, 1976.

13. Gresser I., Tovey M.G., Maury C. and Bandu M-T.
Role of interferon in the pathogenesis of virus diseases in mice as demonstrated by the use of anti-interferon serum. II. Studies with herpes simplex, Moloney sarcoma, vesicular stomatitis, Newcastle disease and influenza virus.
J. Exp. Med., **144** : 1316-1323, 1976.

14. Gresser I., Maury C. and Tovey M.G.
Interferon and murine leukemia VII. Therapeutic effect of interferon preparations after diagnosis of lymphoma in AKR mice.
Int. J. Cancer, **17** : 647-651, 1976.

15. Stewart II, W.E., Gresser I., Tovey M.G., Bandu M-T. and Le Goff S.
Identification of the cell-multiplication-inhibitory factors in interferon preparations as interferons.
Nature, **262** : 300-302, 1976.

16. Lindahl P., Gresser I., Leary P. and Tovey M.G.
Interferon treatment of mice : enhanced expression of histocompatibility antigens on lymphoid cells.
Proc. Natl. Acad. Sci. USA, **73** : 1284-1287, 1976.

17. Gresser I., Morel-Maroger L., Maury C., Tovey M.G. and Pontillon F.
Progressive glomerulonephritis in mice treated with interferon preparations at birth.
Nature, **263** : 420-422, 1976.

18. Lindahl P., Gresser I., Leary P. and Tovey M.G.
Enhanced expression of histocompatibility antigens of lymphoid cells in mice treated with interferon.
J. Inf. Dis., supplement **133** : A66-A68, 1976.

19. Tovey M.G., Begon-Lours J., Gresser I. and Morris A.G.

Marked enhancement of interferon production in 5-bromodeoxyuridine treated human lymphoblastoid cells.
Nature, **267** : 455-457, 1977.

20. Tovey M.G., Bandu M-T., Begon-Lours J., Brouty-Boyé D. and Gresser I.
    Antiviral activity of bovine interferons on primate cells.
    J. gen. Virol., **36** : 341-344, 1977.

21. Brouty-Boyé D. and Tovey M.G.
    Inhibition of thymidine uptake in chemostat cultures of L1210 cells.
    Intervirology, **9** : 243-252, 1978

22. Gresser I., Maury C. and Tovey M.G.
    Efficacy of combined interferon cyclophosphamide therapy after diagnosis of lymphoma in AKR mice.
    Eur. J. Cancer, **14** : 97-99, 1978.

23. Gresser I., Maury C., Bandu M-T., Tovey M.G. and Maunoury M-T.
    Role of endogenous interferon in the antitumor effect of poly I.C, and statolon as demonstrated by the use of anti-mouse interferon serum.
    Int. J. Cancer, **21** : 72-77, 1978.

24. Rivière Y., Gresser I., Guillon J-C. and Tovey M.G.
    Inhibition by anti-interferon serum of lymphocytic choriomeningitis virus disease in suckling mice.
    Proc. Natl. Acad. Sci. USA, **74** : 2135-2138,1978.

25. De Maeyer-Guignard J., Tovey M.G., Gresser I. and De Maeyer E.
    Purification of mouse interferon by sequential affinity chromatography on poly(U) and antibody-agarose columns.
    Nature, **271** : 622-625, 1978.

26. Gresser, I. and Tovey, M.G.
    Antitumor effects of interferon.
    Biochem. Biophys. Acta, **516** : 231-247, 1978.

27. Tovey M.G., Lenoir G. and Begon-Lours J.
    Activation of latent Epstein-Barr virus by antibody to human IgM.
    Nature, **276** : 270-272, 1978.

28. Gresser I., Tovey M.G., Maury C. and Bandu M-T.
    Role of interferon in the pathogenesis of herpes simplex virus disease in mice.
    In: Oncogenesis and Herpes Viruses, vol. 3 (eds. De Thé G., Henle W. & Rapp F.) (Int. Agency Res. Cancer, Lyon) pp. 1049-1054, 1979.

29. Tovey M.G. and Brouty-Boyé D.

The use of the chemostat to study the relationship between cell growth rate, viability and the effect of interferon on L1210 cells.
Exp. Cell Res., **118** : 383-388, 1979.

30. Gresser I., Morel-Maroger L., Châtelet F., Maury C., Tovey M.G., Bandu M-T., Buywid J. and Delauche M.
Delay in growth and the development of nephritis in rats treated with interferon preparations in the neonatal period.
Am. J. Pathol., **95** : 329-346, 1979.

31. Tovey M.G., Lenoir G., Begon-Lours J., Tapiero H. and Rochette-Egly C.
The effect of mitogens on the expression of Epstein-Barr virus antigens in human lymphoid cell lines.
J. Immunol., **123** : 138-142, 1979.

32. Tovey M.G., Rochette-Egly C. and Castagna M.
Effect of interferon on concentrations of cyclic nucleotides in cultured cells.
Proc. Natl. Acad. Sci. USA, **76** : 3890-3893, 1979.

33. Gresser I., De Maeyer-Guignard J., Tovey M.G. and De Maeyer E.
Electrophoretically pure mouse interferon exerts multiple biologic effects.
Proc. Natl. Acad. Sci. USA, **76** : 5308-5312, 1979.

34. Gresser I., Morel-Maroger L., Rivière Y., Guillon J-C., Tovey M.G., Woodrow D., Sloper J.C. and Moss J.
Interferon induced disease in mice and rats.
Annals New York Acad. Sci., **350** : 12-20, 1980.

35. Tovey M.G. and Rochette-Egly C.
The effect of interferon on cyclic nucleotides.
Annuals New York Acad. Sci., **350** : 266-278, 1980.

36. Lenoir G., Tovey M.G. and Lavoué M-F.
Induction of Epstein-Barr virus early antigen by phytohaemagglutinin in the presence of 5-iodo-2'-deoxyuridine : application to EBV serology.
J. Immunol. Methods, **34** : 23-29, 1980.

37. Tovey M.G., Rochette-Egly C. and Castagna M.
Correlation between growth rate, cell density, and intracellular concentrations of cyclic nucleotides in chemostat cultures of mouse L1210 cells.
J. Cell Physiol., **105** : 363-367, 1980.

38. Tovey M.G.
The cultivation of animal cells in the chemostat : application to the study of tumor cell multiplication.

In: Advances in Cancer Research, **33** : 1-37, 1980.

39. Adam C., Thoua Y., Ronco P., Verroust P., Tovey M.G. and Morel-Maroger L.
The effect of exogenous interferon : acceleration of autoimmune and renal diseases in (NZB/W)F1 mice.
Clin. Exp. Immunol., **40** : 373-382, 1980.

40. Tovey M.G.
Viral latency and its importance in human disease.
Path. Biol. **26** : 631-634, 1980.

41. Gresser I., Morel-Maroger L., Rivière Y., Guillon J-C., Tovey M.G. and Woodrow D.
Interferon-induced disease in mice and rats.
In: Communications of liver cells. Falk Symposium 27 - MTP Press Ltd. (H. Hopper et al., eds) 1980.

42. De Maeyer-Guignard J., De Maeyer E., Tovey M.G. and Gresser I.
Electrophoretically pure mouse interferon exerts multiple biological effects.
Annuals New York Acad. Sci., **350** : 347-353, 1980.

43. Tovey M.G.
Use of the chemostat culture for study of the effect of interferon on tumor cell multiplication.
In : Methods in Enzymology, vol. 79 : Interferons Part B (S. Pestka, ed.) Academic Press, New York, pp. 391-404, 1981.

44. Tovey M.G. and Rochette-Egly C.
Rapid increase in guanosine 3'5'-monophosphate in interferon treated cells.
In: The Biology of the Interferon System (E. De Maeyer and H. Schellekens, eds.), Elsevier/North Holland, pp. 165-168, 1981.

45. Tovey M.G. and Rochette-Egly C.
Rapid increase in guanosine 3'5'-monophosphate in interferon treated mouse leukemia L1210 cells : relationship to the development of the antiviral state and inhibition of cell multiplication.
Virology, **115** : 272-281, 1981.

46. Tovey M.G., Dron M. and Gresser I.
Interferon enhances the expression of Epstein-Barr virus early antigen in Daudi cells.
J. gen. Virol., **60** : 31-38, 1982.

47. Yerushalmi A., Tovey M.G. and Gresser I.
    Antitumor effect of combined interferon and hyperthermia in mice.
    Proc. Soc. Exp. Biol. Med., **169** : 413-415, 1982.

48. Kawade Y., Aguet M. and Tovey M.G.
    Antigenic correlations between components of C243 and L cell interferons.
    Antiviral Res., **2** : 155-159, 1982.

49. Keller R., Aguet M., Tovey M.G. and Stitz L.
    Prevention of interferon-induced augmentation of cellular antitumor effector mechanisms by phorbol esters.
    Cancer Res., **42** : 1468-1472, 1982.

50. Tovey M.G., Gresser I., Rochette-Egly C., Begon-Lours-Guymarho J., Bandu M-T., Maury C.
    Indomethacin and aspirin do not inhibit the antiviral and anti-proliferative action of interferon.
    J. gen. Virol., **63** : 505-508, 1982.

51. Tovey M.G. and Gresser I.
    Interferon-induced disease in mice and rats.
    In: The Clinical Potentials of Interferons in Viral Diseases and Malignant Tumors. (R. Kono and J. Vilcek, eds.) University of Tokyo Press, pp. 311-319, 1982.

52. Tovey M.G.
    Interferon and cyclic nucleotides.
    In: Interferon 4, 1982 (I. Gresser, ed.) Academic Press, London-New York, pp. 23-46.

53. Tovey M.G., Rochette-Egly C. and Kolb J-P.
    Interferon enhancement of natural killer cell cytotoxicity : role of cyclic nucleotides.
    J. Interferon Res., **2** : 549-561, 1982.

54. Rochette-Egly C. and Tovey M.G.
    Interferon enhances guanylate cyclase activity in human lymphoma cells.
    Biochem. Biophys. Res. Commun., **107** : 150-156,1982.

55. Rosa F., Fellous M., Dron M., Tovey M.G. and Revel M.
    Presence of an abnormal β2-microglobulin in mRNA in Daudi cells : induction by interferon.
    Immunogenetics, **17** : 125-131, 1983.

56. Tovey M.G., Vincent C., Gresser I. and Revel M.
    The effect of DNA methylation on the expression of human interferon α genes.
    In: The Biology of the Interferon System. Proceedings of the International Meeting on the Biology of the Interferon System, Rotterdam. (Eds. E. De Maeyer, G. Galasso and H. Schellekens)Elsevier/North Holland, pp. 45-50, 1983.

57. Hoshino A., Takenaka H., Mizukoshi O., Imanishi J., Kishida T. and Tovey M.G.
Effect of anti-interferon serum of influenza virus infection in mice.
Antiviral Res., **3** : 59-65, 1983.

58. Tovey M.G.
The cultivation of animal cells in continuous-flow culture.
In: Animal Cell Biotechnology (eds R.E. Spier and B. Griffiths) Academic Press, New York, vol.**1**, pp. 195-210, 1985.

59. Dron M. and Tovey M.G.
Isolation of Daudi cells with reduced sensitivity to interferon. I. Characterization.
J. gen. Virol., **64** : 2641-2647, 1983.

60. Tovey M.G., Dron M., Mogensen K.E., Lebleu B., Mechti N., and Bégon-Lours-Guymarho J.
Isolation of Daudi cells with reduced sensitivity to interferon. II. On the mechanisms of resistance.
J. gen. Virol., **64** : 2649-2653,1983.

61. Gresser I., Belardelli F., Maury C., Maunoury M-T. and Tovey M.G.
Injection of mice with antibody to interferon enhances the growth of transplantable murine tumors.
J. Exp. Med., **158** : 2095-2107, 1983.

62. Tovey M.G.
Interferon : immunomodulator and antitumor agent.
In: Immunomodulation. (ed. H. Hugh Fudenberg, H.D. Whitten and F. Ambrogi) Plenum Publishing Corporation, pp. 155-159, 1984.

63. Souyri-Caporale M., Tovey M.G., Ono K., Jasmin C. and Chermann J-C.
Modulation by the polyoxotungstate HPA-23 of early antigen expression in Raji cells treated with iododeoxyuridine.
J. gen. Virol., **65** : 831-835, 1984.

64. Rochette-Egly M. and Tovey M.G.
Natural killer cell cytotoxicity : role of calmodulin.
Biochem. Biophys. Res. Commun., **121** : 478-486, 1984.

65. Tovey M.G. and Gresser I.
Interferon induced disease.
In: "The Physiology and Pathology of Interferon System (ed. L. Borecky and V. Lackovic). Contr. Oncol., vol. **20**, pp. 196-204, Karger, Basel, 1984.

66. Pozzetto B., Mogensen K.E., Tovey M.G. and Gresser I.

Characterization of auto-antibodies to human interferon in a patient with a varicella zoster.
J. Infect. Dis., **105** : 707-713, 1984.

67. Gresser I., Belardelli F., Vignaux F., Maury C., and Tovey M.G.
Role of endogenous "spontaneous" interferon in mice.
In: "The Biology of the Interferon System, 1984", Heidelberg. H. Kirchner and H. Schellekens (eds.). Elsevier Science Publishers B.V., Amsterdam, The Netherlands, p. 16, 1984.

68. Dron M. and Tovey M.G.
Interferon resistant Daudi cells : on the mechanisms of resistance.
In: "The Biology of the Interferon System, 1984", Heidelberg. H. Kirchner and H. Schellekens (eds.). Elsevier Science Publishers B.V., Amsterdam, The Netherlands, p. 40, 1984.

69. Rochette-Egly C. and Tovey M.G.
Cyclic GMP levels in interferon treated cells.
Antiviral Res., **5** : 127-135, 1985.

70. Gresser I., Vignaux F., Belardelli F., Tovey M.G. and Maunoury M-T.
Injection of mice with antibody to mouse interferon α/β decreases the level of 2'-5'oligoadenylate synthetase in peritoneal macrophages.
J. Virol., **53** : 221-227, 1985.

71. Dron M., Tovey M.G. and Eid P.
Interferon induced proteins : relationship to the phenotype of interferon resistance in cloned Daudi cells.
J. gen. Virol., **66** : 785-795, 1985.

72. Gresser I., Belardelli F., Vignaux F. and Tovey M.G.
Is interferon produced constitutively in normal mice ?
In: The Interferon System. (Eds. F. Dianzani and G.B. Rossi), vol. **24**, pp. 99-102, Raven Press, New York, 1985.

73. Le Roscouet D., Vodjdani G., Lemaigre-Dubreuil Y., Tovey M.G., Latta M. and Doly J.
Structure of a murine alpha interferon pseudogene with a repetitive R-type sequence in the 3' flanking region.
Mol. Cell. Biol., **5** : 1343-1348, 1985.

74. Tovey M.G. and Gresser I.
Detection of interferon messenger RNA in the organs of normal mice.
In: The Interferon System. (Eds. F. Dianzani and G.B. Rossi), vol. **24**, pp. 109-113, Raven Press, New York, 1985.

75. Gresser I., Belardelli F., Vignaux F., Maury C. and Tovey M.G.
Role of endogenous "spontaneous" interferon in mice.

In: The Biology of the Interferon System. (Eds. H. Kirchner and H. Schellekens), pp. 157-160, Elsevier Publishers, B.V., Amsterdam, The Netherlands, 1986.

76. Dron M., Tovey M.G. and Uzé G.
Isolation of Daudi cells with reduced sensitivity to interferon. IV. Characterization of clones with altered binding of human interferon subspecies.
J. gen. Virol., **67** : 663-669, 1986.

77. Dron M., Modjtahedi N., Brison O. and Tovey M.G.
Interferon modulation of c-myc expression in cloned Daudi cells : relationship to the phenotype of interferon resistance.
Mol. Cell. Biol., **6** : 1374-1378, 1986.

78. Gresser I., Belardelli F., Maury C., Tovey M.G. and Maunoury M-T.
Antitumor effects of interferon in mice injected with interferon-resistant Friend leukemia cells. IV. Definition of optimal treatment regimens.
Int. J. Cancer, **38** : 251-257, 1986.

79. Meister A., Uzé G., Mogensen K.E., Gresser I., Tovey M.G., Grütter M. and Meyer F.
Biologic activities and receptor binding of 2 human recombinant interferons and their hybrids.
J. gen. Virol., **67** : 1633-1643, 1986.

80. Tovey M.G., Streuli M., Gresser I., Gugenheim J., Blanchard B., Guymarho J., Vignaux F. and Gigou M.
Interferon messenger RNA is produced constitutively in the organs of normal individuals.
Proc. Natl. Acad. Sci. USA, **84** : 5038-5042, 1987.

81. Tovey M.G., Gresser,I.
Interferon β-2 is expressed at high levels in the organs of normal individuals.
J. Interferon Res. **7** : 701, 1987.

82. Palmieri M., Tovey M.G.
Evidence for the presence of regulatory proteins in the 5'untranslated region of the interferon α-1 gene in cells from normal individuals.
J. Interferon Res**7** : 711, 1987.

83. Besançon F., Silbermann F., Dron M., Tovey M.G., Thang M.N., and Bourgeade M-F.
Relationship between inhibition of cell growth and of transferrin receptor expression by interferon (IFN) α : Studies in IFN-sensitive and IFN-resistant Daudi cells.
J. gen. Virol., **68** : 2647-2654, 1987.

84. Tovey M.G., and Gresser I.
Interferon as an antiviral agent : role of endogenous interferon.

In : Development in Antiviral Chemotherapy (ed. M. Alexander). Ecomed Landsberg, 1987.

85. Tovey M.G.
The expression of cytokines in the organs of normal individuals : role in homeostasis.
J. of Biol. Reg. & Homeost. Agents, **2** : 87-92, 1988.

86. Tovey M.G., Content J., Gresser I., Gugenheim J., Blanchard, B., Poupart, P., Gigou, M., Shaw, A., and Fiers, W.
Genes for interferon β-2 (interleukin-6), tumor necrosis factor, and interleukin-1 are expressed at high levels in the organs of normal individuals.
J. Immunology, **141** : 3106-3110, 1988.

87. Gresser, I., Maury, C., Vignaux, F., Haller, O., Belardelli, F. and Tovey, M.G.
Antibody to mouse interferon α/β abrogates resistance to the multiplication of Friend erythroleukemia cells in the livers of allogeneic mice.
J. Exp. Med., **168** : 1271-1291, 1988.

88. Tovey, M.G. and Palmieri, M.
Expression of interferon and cytokine genes in normal individuals and in patients with auto-immune diseases. The 1988 ISIR meeting on Interferon and Cytokines, Kyoto 14-18, 1988.
J. of Interferon Res. **8** suppl.1 : S156, 1988.

89. Dron, M., Tovey, M.G., Maury, C., Eid, P., Meyer, F. and Gresser, I.
Production and in vivo biologic actions of recombinant mouse interferon α2.
J. Biol. Regul. & Homeost. Agents, **3** : 13-19, 1989.

90. Tovey, M.G.
Expression of the genes of interferons and other cytokines in normal and diseased tissues of man.
Experientia, **45** : 526-535, 1989.

91. Tovey, M.G., Gresser, I., Blanchard, B. and Guymarho, J.
Expression of IL-6 in normal individuals and in patients with autoimmune disease.
In "Regulation of the acute phase and immune responses: Interleukin-6.
Ann. N.Y. Acad., **557** : 363-373, 1989.

92. Tovey, M.G., and Palmieri, M.
Expression of interferon and cytokine genes in normal individuals and in patients with autoimmune diseases.
In "The Biology of the Interferon System 1988". Kawade, Y., and Kobayashi, eds., pp. 329-334, 1989.

93. Dron, M., Lacasa, M., Tovey, M.G.

Priming affects the activity of a specific region of the promoter of the human beta IFN gene.
Mol. & Cell. Biol., **10** : 854-858, 1990.

94. Palmieri, M., and Tovey, M.G.
Genomic footprinting: Detection of putative regulatory proteins in the promoter region of the interferon α-1 gene in normal human tissues.
Mol. & Cell. Biol., **10** : 2554-2561, 1990.

95. Vanden Broecke, C., Caillat-Zucman, S., Legendre, C., Noeil, L-H., Kreis, H., Woodrow, D., Bach, J-F., and Tovey, M.G.
Differential in situ expression of cytokines in renal allograft rejection.
Transplantation, **51** : 602-609, 1991.

96. Caillat-Zucman, S., Vanden Broecke, C., Legendre, C., Noel, L-H., Kreis, H., Bach, J-F., and Tovey, M.G.
Differential in situ expression of cytokine genes in human renal rejection.
Transplantation Proceedings, **23/1** : 229-231, 1991.

97. Vanden Broecke, C., and Tovey, M.G.
The expression of the genes of class I interferons and interleukin 6 in individual cells.
J. Interferon Res., **11** : 91-1031991.

98. Vanden Broecke, C., Caillat-Zucman, S., Legendre, C., Noel, L-H., Kreis, H., Bach, J.-F., Tovey, M.G.
Study of graft rejection using molecular biology technics.
In : "Immunologic, metabolic, and infectious aspects of hepatic transplantation", ed. D.A. Vuitton, John Libbey LTD, London, 1991.

99. Tovey, M.G.
The antiproliferative action of interferons.
Seminars in Oncology, W.B. Saunders Company, 1991.

100. Tovey, M.G., Gugenheim, J., Guymarho, J., Blanchard, B., Vanden Broecke, C., Gresser, I., Bismuth, H., and Reynes, M.
Genes for interleukin-1, interleukin-6, and tumor necrosis factor are expressed at markedly reduced levels in the livers of patients with severe liver disease.
Autoimmunity, **10 /4**: 297-310, 1991.

101. Proietti, E., Vanden Broecke, C., Di Marzio, P., Gessani, S., Gresser, I., and Tovey, M.G.
Specific interferon genes are expressed in individual cells in the peritoneum and bone marrow of normal mice.
J. Interferon Res., **12** : 27-34, 1992.

102. Dron, M., and Tovey, M.G.
Interferon α/β gene structure and regulation.
In : Interferon: Principles and Medical Applications, (eds. S. Baron *et al.*), 33-45, 1992.

103. Dron, M., Rebouillat, D., and Tovey, M.G.
Tandem repeat polymers of a critical region of the human interferon β promoter exhibit a marked constitutive activity and enhanced responsiveness to transcriptional regulators in transfected HeLa cells.
J. Interferon Res., **12** : 377-384, 1992.

104. Tovey, M.G., Lebon, P., Rizza, P., and Lallemand, C.
The expression of interferon genes in the tissues of AIDS-patients.
In : The IFN-system in HIV-infection", VII European Interferon Workshop, 26-28 February, 1992, Hanover.

105. Tovey, M.G., Lebon, P., Meyer, F., Hurtrel, B., Gresser, I., Venet, A., and Girard, M.
The role of interferon in the development of AIDS;
"1992 Annual Meeting of the International Society for Interferon Research" September 28- October 2, 1992, Toronto.

106. Lallemand, C., and Tovey, M.G.
Differential expression of IFN genes in cells infected with HIV.
"1992 Annual Meeting of the International Society for Interferon Research" September 28- October 2, 1992, Toronto.

107. Krivine, A., Tovey, M.G., Taty-Taty, R., and Lebon, P.
Endogenous interferon alpha in newborns from HIV seropositive mothers.
"1992 Annual Meeting of the International Society for Interferon Research" September 28- October 2, 1992, Toronto.

108. Tovey, M.G., Lebon, P., Meyer, F., Hurtrel, B., Gresser, I., Venet, A., Girard, M., and Aubertin, A.-M.
The role of interferon in the development of AIDS.
"Tenth Annual Symposium on Nonhuman Primate Models for AIDSé, San Juan, Puerto Rico, 14-22 November, 1992.

109. Gugenheim, J., Tovey, M.G., Gigou, M., Crafa, F., Fabiani, B., Reynes, M., Bismuth, H.
Prolongation of hert allograft survival in rats by anti-interferon antibodies and low-dose cyclosporine.
Transplant. International, 1992.

110. Benoit, P., Maguire, D., Plavec, I., Kocher, H-P., Tovey, M.G., and Meyer, F.

A monoclonal antibody to recombinant human IFN α receptor inhibits biological activity of several species of human IFN α, IFN β and IFN ω.
J. Immunol., **150** : 707-716, 1993.

111. Tovey, M.G., Deglise-Favre, A., and Schoevaert, D.
Differential in situ expression of cytokine genes in human renal transplantation.
Kidney International, **43** : 129, 1993.

112. Dron, M. and Tovey, M.G.
Interferon-resistant Daudi cells are deficient in interferon α-induced ISGF3 α production, but remain sensitive to the interferon α-induced increase in ISGF3 γ content.
J. Interferon Res., **13** : 377-383, 1993.

113. Tovey, M.G.
Le Patrimoine immatériel et intellectuel : Le séquençage du génome humain.
Actes du Forum d'Iéna, **1** : 76-78, 1993.

114. Tovey, M.G., Lebon, P., Meyer, F., Hurtrel, B., Gresser, I., Venet, A., Girard, M., Aubertin, A.M.
Antibody to the human IFN α receptor reduces the loss of CD4+ T cells in macaques infected with the simian immunodeficiency virus (SIV).
J. Interferon Res. **13** (suppl. 1) : S202, 1993.

115. Tovey, M.G.
Differences between IFN-α and IFN-β both from the point of view of gene regulation and physico-chemical properties etc.
In : Proceedings of 1993 International Symposium on Viral Hepatitis and Liver Diseases.
Medical Tribune, Inc.,1994.

116. Militerno, G., Gugenheim, J., Cuomo, O., Hofman, P., Mouiel, J., Tovey, M.G.
Synergistic interaction between anti-IFN α/β antibody and low doses of cyclosporin therapy prolongs heart transplants in rats.
Transplantation Proc., **26** : 3050-3051, 1994.

117. Tovey, M.G., Lebon, P., Eid, P., Meyer, F., Hurtrel, B., Gresser, I., Venet, A., Girard, M. and Aubertin, A-M.
Antibody to the human IFN α receptor reduces the loss of CD4+ T cells in macaques infected with the simian immunodeficiency virus (SIV).
BioTech RIA '94 Congress, Florence, April 11-13, 1994.

118. Lallemand, C., Rizza, P., Lebon, P. and Tovey, M.G.
Mechanisms of the inhibition of the interferon response in HIV infected cells.
BioTech RIA '94 Congress, Florence, April 11-13, 1994.

119. Tovey, M.G., Benizri, E., Gugenheim, J., Bernard, G., Eid, P., Hofman, P.

An interferon alpha antagonist induces permanent allograft survival in cynomologus monkeys
J. Interferon Res., **14** : S168, 1994.

120. Lallemand, C., Rizza, P., Lebon, P., Tovey, M.G.
Constitutive expression of specific type I interferons in peripheral blood leukocytes from normal individuals and in the promonocytic cell line U937
J. Interferon Res., **14** : S62, 1994.

121. Abramovich, C., Shulman, L.M., Ratovitski, E., Harroch, S., Tovey, M.G., Eid, P., and Revel, M.
Differential tyrosine phosphorylation of the IFNAR chain of the type I interferon receptor and of an associated surface protein in response to interferon-α and interferon-β
EMBO J., **13** : 5871-5877, 1994.

122. Eid, P. and Tovey, M.G.
Characterization of a domain of a human type I interferon receptor protein involved in ligand binding.
J. Interferon Res., **15** : 205-211, 1995.

123. Gresser, I., Maury, C., Kaido, T., Bandu, M-T., Tovey, M.G., Maunoury, M-T., Fantuzzi, L., Gessani, S., Greco, G., Proietti, E., and Belardelli, F.
The essential role of endogenous IFN α/β in the antimetastatic action of sensitized T lymphocytes in mice injected with Friend erythroleukemia cells.
Int. J. Cancer, **63** : 726-731, 1995.

124. Tovey, M.G., Benizri, E., Gugenheim, J., Bernard, G., Eid, P., Blanchard, B., and Hofman, P.
Role of the type I interferons in allograft rejection.
J. Leukocyte Biol., **59** : 512-517, 1996.

125. Weill, D., Gay, F., Tovey, M.G., and Chouaib, S.
Induction of tumor necrosis factor-α expression in human T lymphocytes following ionizing gamma irradiation.
J. Interferon & Cytokine Res., **16** : 395-402, 1996.

126. Khatissian, E., Tovey, M.G., Cumont, M-C., Monceaux, V., Lebon, P., Montagnier, L., Hurtrel, B., and Chakrabarti, L.
Rapid interferon-alpha response does not prevent the establishment of a high viral burden in primary SIV infection.
AIDS Res. & Human Retroviruses, **12** : 1273-1278, 1996.

127. Merola, M., Blanchard, B., and Tovey, M.G.
    The κB enhancer of the human interleukin-6 promoter is the unique IL-1β and TNF α responsive element/requirement for members of the C/EBP family for activity.
    J. Interferon & Cytokine Res., **16** : 783-798, 1996.

128. Lallemand, C., Lebon, P., Rizza, P., Blanchard, B., and Tovey, M.G.
    Constitutive expression of specific interferon isotypes in peripheral blood leukocytes from normal individuals and in promonocytic U937 cells.
    J. Leukocyte Biol., **60** : 137-146, 1996.

129. Cursio, R., Gugenheim, J., Tovey, M., Lasfar, A., Crafa, F., Michiels, J.-F., and Mouiel, J.
    Protective properties of anti-IFN α/β antibodies in normothermic hepatic ischaemia in the rat.
    Transplantation Proc., **28** : 73-74, 1996.

130. Lang, M-C., Cansier, C., Rizza, P., Jasmin, C., Tovey, M.G., and Eid, P.
    A comparative study of the expression of cellular proteins in uninfected and HIV infected U937 cells using two dimensional SDS polyacrylamide gel electrophoresis.
    Chemico-Biological Interactions, **103** : 179-186, 1997.

131. Lallemand, C., Bayat-Sarmadi, M., Blanchard, B., and Tovey, M.G.
    Identification of a novel transcriptional regulatory element common to the P53 and IRF1 genes.
    The J. Biol. Chemistry, **272/47** : 29801-29809, 1997.

132. Benizri, E., Gugenheim, J., Lasfar, A., Eid, P., Blanchard, B., Lallemand, C., and Tovey, M.G.
    Prolongation allograft survival in cynomologus monkeys treated with a monoclonal antibody human type I interferon receptor and low doses of cyclosporin A.
    J. Interferon & Cytokine Res., **18** : 273-284, 1998.

133. Dandoy-Dron, F., Guillo, F., Benboudjema, L., Deslys, J-Ph., Lasmesas, C., Dormont, D., Tovey, M.G., and Dron, M.
    Gene expression in scrapie : cloning of a new scrapie responsive gene and the identification of increased levels of seven other mRNA transcripts.
    J. Biol. Chem., **273** : 7691-7697, 1998.

134. Dron, M., Dandoy-Dron, F., Guillo, F., Benboudjema, L., Hauw, J.-J., Lebon, P., Dormont, D, and Tovey, M.G.
    Characterization of the human analogue of a Scrapie responsive gene.
    J. Biol. Chem., **273** : 18015-18018, 1998.

135. Armenante, F., Merola, M., Furia, A., Tovey, M.G., and Palmieri, M.
    Interleukin-6 repression is associated with a distinctive chromatin structure of the gene.
    Nucleic Acids Res., **27** : 4483-4490, 1999.

136. Dron, M., Hameau, L., Benboudjema, L., Guymarho, J., Cajean-Feroldi, C., Rizza, P., Godard, C., Tovey, M.G., Jasmin, C., and Lang, M.-C.
Cloning of a long HIV-1 readthrough transcript and detection of an increased level of Egr-1 mRNA in chronically infected U937.
Arch. Virol., **144** : 19-28, 1999.

137. Eid, P., Méritet, J.-F., Maury, C., Lasfar, A., Weill, D. and Tovey, M.G.
Oromucosal interferon therapy : Pharmacokinetics and pharmacodynamics.
J. Interferon & Cytokine Res., **19** : 157-169, 1999.

138. Tovey, M.G. and Maury, C.
Oromucosal interferon therapy : Marked antiviral and antitumor activity.
J. Interferon & Cytokine Res., **19** : 145-155, 1999.

139. Tovey, M.G., Méritet, J-F., Guymarho, J. and Maury, C.
Mucosal cytokine therapy: Marked antiviral and antitumor activity.
J. Interferon & Cytokine Res., **19** : 911-921, 1999.

140. Palmieri, M., Sasso, M.P., Monese, R., Merola, M., Faggioli, L., Tovey, M., and Furia, A.
Interaction of the nuclear protein CBF1 with the kB site of the IL-6 gene promoter.
Nucleic Acids Res. **27** : 2785-2791, 1999.

141. Dandoy-Dron, F., Benboudjema, L., Guillo, F., Jaegly, A., Jasmin, C., Dormont, D., Tovey, M.G., and Dron, M.
Enhanced levels of scrapie responsive gene mRNA in BSE-infected mouse brain.
Mol. Brain Res., **76** : 173-179, 2000.

142. Eid, P., Langer, J.A., Bailly, G., Lejealle, R., Guymarho, J., and Tovey, M.G.
Localization of a receptor nonapeptide with a possible role in the binding of the type I interferons.
Eur. Cytokine Network, **11** : 560-573, 2000.

143. Dron, M., Tartare, X., Guillo, F., Haik, S., Barbin, G., Maury, C., Tovey, M.G., and Dandoy-Dron, F.
Mouse scrapie responsive gene 1 (Scrg1): Genomic organization, physical linkage to Sap30, genetic mapping on chromosome 8, and expression in neuronal primary cell cultures.
Genomics, **70** : 140-149, 2000.

144. Tovey, M.G.
Guide to novel therapeutic targets just misses target.
Trends in Immunol., **22** : 59-60, 2001.

145. Manca, C., Tsenova, L., Bergold, A., Freeman, S., Tovey, M.G., Musser, J.M., Barry III, C.E., Freedman, V.H., and Kaplan, G.
Virulence of a Mycobacterium tuberculosis clinical isolate in mice is determined by failure induce Th1 type immunity and is associated with induction of IFN α–β.
Proc. Natl. Acad. Sci. USA, **98** : 5752-5757, 2001.

146. Sonnenfeld, G., Tovey, M.G., Schellekens, H., Kinney, K.S., Belay, T., Morton, D.S., Austin, C.A., Reitman, M., Fong, A.T., and Vaughan, H.S.
Efficacy and safety of oral/sublingual, intranasal and intraperitoneal ecombinant murine interferon in the treatment of murine encephalomyocarditis virus.
J. Interferon Cytokine Res., **21** : 539-545, 2001.

147. Schellekens, H., Geelen, G., Méritet, J-F., Maury, C., and Tovey, M.G.
Oromucosal interferon therapy : Relationship between antiviral activity and virus load.
J. Interferon Cytokine Res., **21** : 575-581, 2001.

148. Méritet, J-F., Maury, C., and Tovey, M.G.
Effect of oromucosal administration of interferon alpha on allergic sensitization and the hypersensitive inflammatory response in animals sensitized to ragween pollen.
J. Interferon Cytokine Res., **21** : 583-593, 2001.

149. Méritet, J-F., Maury, C., and Tovey, M.G.
Induction of tolerance to recombinant therapeutic proteins.
J. Interferon Cytokine Res., **21** : 1031-1038, 2001.

150. Lallemand C., Palmieri, M., Blanchard, B., Méritet, J-F., and Tovey, M.G.
GAAP-1 : A transcriptional activator of p53 and IRF-1 possesses pro-apoptotic activity.
EMBO Reports, **3** : 153-158, 2002.

151. Dron, M., Méritet, J-F., Dandoy-Dron, F., Meyniel, J-Ph., Maury, C., Tovey, M.G.
Molecular cloning of *ADIR*, a novel interferon responsive gene encoding a protein related to the torsins.
Genomics, **79** : 315-325, 2002.

152. Giron-Michel, J., Weill, D., Bailly, G., Legras, S., Nardeux, P-C., Azzarone, B., Tovey, M.G., Eid, P.
Direct signal transduction via functional interferon □□□ receptors in CD34+ hematopoietic stem cells.
Leukemia, **16** : 1135-1142, 2002.

153. Moreira, A.L., Tsenova, L., Aman M.H., Bekker, L-G., Freeman, S., Mangaliso, B., Sshröder, U., Jagirdar, J., Rom, W.N., Tovey, M.G., Freedman, V.H., and Kaplan, G.
Mycobacterial antigens exacerbate disease manifestations in *Mycobacterium tuberculosis*-infected mice.
Inf. and Immun., **70** : 2100-2107, 2002.

154. Tovey, M.G.
Oromucosal cytokine interactions.
J. Interferon & Cytokine Res., **22** :163-164, 2002.

155. Guinet, E., Mimaud, V., Rossignol, P., Hameau, L., Benboudgema, L., Jasmin, C., Tovey, M.G., Bélec, L., Lang, M-C.
Do retroviruses preferentially integrate within highly plastic regrions of the human genome ?
Medical Hypotheses**, 60** : 293-297, 2003.

156. Dandoy-Dron, F., Griffond, B., Mishal, Z., Tovey, M.G., Dron, M.
Scrg-1, a novel protein of the CNS is targeted to the large dense-core vesicles in neuronal cells.
Eur. J. Neurosciences, **18** : 2449-2459, 2003.

157. Cajean-Feroldi, C., Nosal, F., Nardeux, P.C., Gallet, X., Guymarho, J., Baychelier, F., Sempé, P., Tovey, M.G., and Eid, P.
Identification of residues of the IFNAR 1 chain of the type I human interferon receptor critical for ligand binding and biological activity.
Biochemistry, **43** : 12498-12512, 2004.

158. Hadj-Slimane, R., Chelbi-Alix, M.K., Tovey, M.G., Bobé, P.
An essential role for IFN-α in the overexpression of Fas ligand on MRL/*lpr* lymphocytes, and on their spontaneous Fas-mediated cytotoxic potential.
J. IFN Cytokines Res**.**, **24** : 717-728, 2004.

159. Porta, C., Hadj-Slimane, R., Nejmeddine, M., Pampin, M., Tovey, M.G., Espert, L., Alvarez, S., Chelbi-Alix, M.K.
Interferons-α and γ induce p53-dependent and –independent apoptosis, respectively
Oncogene, **24** : 605-615, 2005.

160. Manca, C., Tsenova, L., Freeman, S., Mangaliso, B., Barczak, A.K., Tovey, M.G., Barry III, C., Kaplan, G.
Induction of type I IFN by the W/Beijing strains is associated with the inhibition of pro-inflammatory cytokine production and reduced Th1 immunity.
J. Interferon & Cytokine Res., **25** : 694-701, 2005.

161. Dron, M., Bailly, Y., Beringue, V., Haeberlé, A-M., Griffond, B., Risold, P-Y., Tovey, M.G., Laude, H., and Dandoy-Dron, F.
Scrg1 is induced in TSE and brain injuries, and associated with autophagy.
Eur. J. Neuroscience, **22** : 133-146, 2005.

162. Dron, M., Bailly, Y., Beringue, V., Haeberlé, A-M., Griffond, B., Risold, P-Y., Tovey, M.G., Laude, H., and Dandoy-Dron, F.
Scrg1, a potential marker of autophagy in TSE.

Autophagy, **22** : 133-146, 2005

163. Tovey, M.G., Lallemand, C., Méritet, J-F., Maury, C.
Adjuvant activity of interferon alpha : mechanism(s) of action.
Vaccine, **24** : 246-249, 2006.

165. Dron, M., Bailly, Y., Beringue, V., Haeberlé, A-M., Griffond, B., Risold, P-Y., Tovey, M.G., Laude, H., and Dandoy-Dron, F.
Scrg1, a potential marker of autophagy in TSE.
Addenda Autophagy,**2**:1, 58-60, 2006

166. Young, H.A. and Tovey, M.G.
Inflammatory gut disease : A new target comes into focus.
Proc. Nat. Acad. Sci. USA, **103**:8303-8305, 2006.

167. Dandoy-Dron, F., Bogdanova, A., Beringue, V., Tovey, M.G., Laude, H., Dron, M.
Infection by ME7 prion is not modified in transgenic mice expressing the yeast chaperone Hsp104 in neurons.
Neuroscience Letters, **405**:181-185, 2006.

168. Lallemand, C., Blanchard, B., Palmieri, M., Lebon, P., May, E., Tovey, M.G.
Single stranded RNA viruses inactivate the transcriptional activity of p53 but induce NOXA-dependent apoptosis via post-translational modifications of IRF-1, IRF-3, and CREB.
Oncogene, July 10, 2006.

169. El-Ghoneimi, A., Cursio, R., Schmid-Alliana, A., Tovey, M.G., and Guggenheim, J.
Phentoxifylline inhibits tumor necrosis factor alpha gene transcription following liver ischemia-reperfusion. J. Hepato Pancreato Billary., in press, 2006

170. Hatzfeld, A., Eid, P., Li, M.L., Peiffer, I., Monier, M-N., Barbet, R., Fortunel, N., Haydont, V., Charbord, P., Tovey, M.G. Hatzfeld, J.
A Subpopulation of high potential-quiescent human mesenchymal stem cells under the control of interferon alpha/bata and TGF-beta.
Leukemia., in press, 2006

171. Currie, A.J., van der Most, R.G., Broomfield, S.A., Prosser, A., Tovey, M.G., Robinson, B.
Eradication of established solid tumors by local reactivation of pre-existing CD8 T cell responses with IFN-a/b-inducing TLR liogands.
J. Immunol., in press, 2007

172. Baychelier, F., Nardeux, P-C, Cajean-Feroldi, C., Ermonval, M., Guymarho, J., Tovey, M.G. Eid, P
Involvement of Gab2 scaffolding adaptor in type I interferon signaling.
Cellular Signalling, in press, 2007

173. Palmer, P., Tovey, M.G., Raschilas, F., Brassart, L., Meritet, J-f., Procher, R., Lebon, P.

Type I interferon sub-types produced by peripheral mononuclear cells from one normal donor stimulated by viral and non-viral factors.
European Cytokine Network in press, 2007